# Effect of Ascorbic Acid, 8-Hydroxyquinoline Sulfate and Sucrose on the Longevity and Anthocyanin Content of Cut Gerbera Flowers

# SAMANEH BANAEE<sup>1</sup>, EBRAHIM HADAVI<sup>1</sup> and PEJMAN MORADI<sup>2</sup>

<sup>1</sup>Department of Horticulture, Karaj branch, Islamic Azad University, Karaj, Iran. <sup>2</sup>Department of Horticulture, Saveh branch, Islamic Azad University, Saveh, Iran.

(Received: April 03, 2013; Accepted: June17, 2013)

## ABSTRACT

The effects of ascorbic acid (AsA), 8-hydroxyquinoline sulfate (8-HQS) and sucrose (Suc) on cut gerbera was studied. AsA (0, 100 and 150 mg L<sup>-1</sup>), 8-HQS (0 and 200 mg L<sup>-1</sup>) and Suc (0 and 30 g L-1) and their combinations were tested as preservative mixtures. Vase life was determined as the days until the flowers started to wilt and then dry weight and anthocyanin content were measured. The following records were analyzed at the 7th day of experiment: mean uptake of preservative solution, dry weight, flower diameter and quality score of cut flowers. The only measure improved by ascorbic acid was the flower diameter. The 8-HQS treatment increased vase life, dry weight, anthocyanin content, fresh weight, flower diameter and mean uptake of preservative solution. Sucrose decreased vase life, anthocyanin content and increased dry weight and flower diameter and mean uptake of preservative solution. The treatment containing the combination of 100 mg L<sup>-1</sup>AsA + 200 mg L<sup>-1</sup>8-HQS + 30 g L<sup>-1</sup> sucrose resulted in the highest vase life but this was not significantly different from the controls. The highest anthocyanin content was noted in the 150 mg L<sup>-1</sup> AsA treatment. The combination of 100 mg L<sup>-1</sup> AsA + 200 mg L<sup>-1</sup>8-HQS + 30 g L<sup>-1</sup> Suc resulted the highest flower diameter. We conclude that AsA could improve the anthocyanin content and flower diameter of flowers and its addition to preservative mixtures based on 8-HQS could improve its effect to a limited extent.

Key words: 8-HQS, dry weight, fresh weight, preservative solution, vase life

Abbreviations: AsA, Ascorbic acid; Suc, Sucrose; 8-HQS, 8-hydroxyquinoline sulfate.

#### INTRODUCTION

Gerbera is a genus of ornamental plants from the Asteraceae family. It has roughly 30 species in the wild, extending to South America, Africa and tropical Asia<sup>1-2</sup>. The inflorescences of *Gerbera hybrida* (Asteraceae) are composed of three different types of flowers (ray, trans, disc) that are tightly packed into a condensed, radially organized capitulum<sup>3</sup>. Most of the present commercially cultivated varieties originate from the artificial crossing progenies of *G. jamesonii* and *G. viridifolia*, both South African species since natural hybrids of the two species have not been found<sup>2</sup>. Gerbera is one of the most important commercially grown flower crops in the world. Gerbera is popular owing to its bright and vivid petal color, which is available in different shades and hues and a large flower size. It has multiple uses in flower arrangements and bouquets and dry flower crafts<sup>4-5</sup>.

However, they often suffer from short vase life<sup>6</sup>. Addition of chemical preservatives to the holding solution is recommended to prolong the vase-life of cut flowers. All holding solutions usually contain two ingredients namely, sugar and germicides. The sugars provide a respiratory substrate, while the germicides control bacterial growth and prevent plugging of the conducting tissues. Therefore, the techniques of prolonging the vase-life of flowers will be a great asset to the growers and users<sup>4</sup>. Different preservative mixtures are used for vaselife prolongation of gerbera including nano silver<sup>6</sup> and silver nitrate+sucrose<sup>4</sup>. Sucrose and ascorbic acid combinations induced extended vase life in Red ginger (*Alpinia purpurata*)<sup>7</sup>. Ascorbic acid at 150 mg l<sup>-1</sup> significantly increased the vase life, fresh weight and percentage of total carbohydrates and increased cut snapdragon flowers vase life<sup>8</sup>. Similar reports with 4 mM AsA on rose exists<sup>9</sup>.

In this study, we aimed to find out the interaction effect between different concentrations of AsA, 8-HQS and Suc on vase life of gerbera cut flowers.

# MATERIALS AND METHODS

#### Plant material and storage conditions

Cut gerbera flowers were obtained from a local commercial greenhouse (Pakdasht, Tehran, Iran), and transported with proper covers immediately to Laboratory (horticulture laboratory of agriculture faculty of Islamic Azad university, Karaj Branch).

Solutions were freshly prepared at the start of experiments. Stems were recut to 45cm length. In this study three levels of ascorbic acid (AsA) (0, 100 and 150 mg l<sup>-1</sup>), two levels of Suc (0 and 3% w/ v) and two levels of 8-HQS (0, 200 mg l<sup>-1</sup>) were applied. After recording the fresh weight, each flower was placed in a bottle containing 400 ml preservative solutions. The flowers were held at ambient temperature (22  $\pm$ 2 !). Vase life, anthocyanin content of petal, fresh weight, flower diameter and uptake of preservative mixture were recorded.

## Vase life

Vase life was determined as the number of days to wilting of flowers. The flowers were checked once a day for signs of deterioration.

#### Determination of anthocyanin content of petal

Anthocyanin was extracted and measured based the method described by Ervin *et al.*,<sup>10</sup>. 0.5 g

petal tissue was extracted with acidified methanol (1% HCl, w/v) for 48 h at room temperature in darkness. The absorbance was measured with spectrophotometer at 530 nm (peak absorption of anthocyanin) and 657 nm (peak absorption of chlorophyll degradation products). The formula  $A^{530}$  – (0.25× $A^{657}$ ) was used to calculate the anthocyanin content.

# Fresh weight and dry weight

All Flowers were weighted at the beginning and at the 7<sup>th</sup> day of experiment. The flowers were dried in oven and the dry weight was calculated.

# Mean uptake of preservative solution per flower in course of experiment

The solution uptake was calculated by subtracting the mean volume of water evaporated from three-control bottles without cut-flowers, from the water decreased in bottles containing flowers during experimental course.

#### Experimental design and statistical analysis

The study was arranged in a factorial test with complete randomized design with three replications. Each replication consisted of three cut flowers. Analysis of variance was performed on the data collected using the general linear model (GLM) procedure of the SPSS software (Version 16, IBM Inc.). The mean separation was conducted by Duncan analysis in the same software (p= 0.05). For the data which were gathered once a day until day 7 of experiment (fresh weight, flower diameter and stem diameter), the GLM-repeated-measures module was applied for factorial analysis and related statistics is presented.

# **RESULTS AND DISCUSSION**

Flower diameter was the only measure that was improved by AsA. Sucrose alone decreased vase life and anthocyanin content and increased dry weight. The flower diameter and mean uptake of preservative solution were affected positively by Suc, conditioned that 8-HQS was present (*Table 1*).

The 8-HQS factor significantly increased the vase life, dry weight, anthocyanin content, fresh

acid (mg/l)	Ascorbic sucrose acid (g/l) (mg/l)	8- hqs (l/gm)	vase life (days)	ife	dry weight (g)	ŧ	fresh weight - day 7 (g)	- <u>1</u> (6)	anthocy anin content (mg/fw)	ç t c	mean uptake of preservativ solution (per flower, ml)	mean uptake of preservative solution (per flower, ml)	flower diameter - day 7 (mm)	stem diamete r - day 7 (mm)	stem diameter r - day 7 (mm)
							(per flc	(per flower, ml)	(Ir						
0	0	0	* 14	# q	2.6	de	26.7	qe	1.34	U	19.0	q	96.8 <sup>de</sup>	7.3	ab
		200	13.4	٩	2.6	de	31.5	bc	1.37	o	38.6	bc	100.2 <sup>cd</sup>	7.5	ab
	30	0	9.6	υ	3.7	υ	26.6	de	0.87	q	9.4	p	94.0 ∘	7.1	ab
		200	17.6	а	5.3	в	37.1	ß	0.87	q	70.3	ß	106.7 <sup>ab</sup>	7.8	а
100	0	0	15.3	ab	2.3	Θ	25.7	٥	1.35	o	12.8	q	94.0 ∘	6.8	Ð
		200	15.6	ab	2.9	q	33.9	ab	1.74	ab	39.1	bc	102.5 bc	7.6	ab
	30	0	8.0	υ	4.0	bc	28.1	cde	0.56	Θ	15.6	p	95.7 <sup>de</sup>	7.2	ab
		200	17.9	а	5.2	в	37.2	Ø	1.44	bc	68.7	ß	108.4 <sup>a</sup>	7.4	ab
150	0	0	15.3	ab	2.8	p	28.8	cde	1.84	ß	21.4	cd	97.9 <sup>cde</sup>	7.5	ab
		200	13.8	p	2.9	p	33.7	ab	1.34	C	49.2	q	102.6 bc	7.7	Ø
	30	0	8.9	υ	3.9	υ	30.9	bcd	0.51	Θ	14.7	p	97.3 <sup>de</sup>	7.1	ab
		200	16.1	ab	4.4	q	35.1	ab	1.51	bc	49.1	q	107.3 <sup>ab</sup>	7.6	ab
Factor significance	nificance														
	AsA		ns §		ns		ns		ns		ns		0.032	ns	
	Suc		0.004		0.000		0.039		0.000		0.049		0.027	ns	
	8-HQS		0.000		0.000		0.000		0.003		0.000		0.000	ns	
	Suc * 8-HQS		0.000		0.001		su		0.001		0.003		0.001	su	

Table 1: The effect of experimental factors and their combinations on observed parameters in gerbera

\* Means in each column followed by similar letters are not significantly different at 5% level. Letters are spaced to highlight the significance. § ns=non significant

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weight, flower diameter and mean uptake of preservative solution. A synergistic effect on dry weight was obvious by interaction of 8-HQS and Suc. The highest dry weight was noted in treatments containing both 30 g L<sup>-1</sup> Suc and 200 mg L<sup>-1</sup> 8-HQS (table 1). The direct effect of 8-HQS on dry weight is significant; being noticeable when AsA is present. When considering the fresh weight the situation is different. The interaction between 8-HQS and Suc is missing while presence of 8-HQS is the determining factor affecting on fresh weight. This phenomena points out the determining effect of 8-HQS on water absorption, as a similar pattern could be observed in effect of 8-HQS on mean uptake of preservative solution. The germistatic effect of 8-HQS that is described elsewhere11-14, and is readily highlighted in our experiment as higher mean uptake of preservative solution values (table 1), where the combinations of 8-HQS and Suc show highest mean uptake of preservative solution values but when Suc is applied alone caused a steep decrease in mean uptake of preservative solution value. This reflects the inhibitory effect of 8-HQS on bacterial colonization and clogging of xylem vessels in cut surface of the flower stem as reported earlier<sup>15-</sup> <sup>16</sup>. The flower diameter followed a similar pattern to mean uptake of preservative solution; being in maximum when both Suc and 8-HQS were present, regardless of AsA concentration.

A synergism between 8-HQS and AsA opposed the decreasing effect of pure Suc on anthocyanin content. The highest anthocyanin content was noted in the treatment containing just 150 mg  $L^{-1}$  AsA.

Contrary to the result obtained on red ginger, snapdragon and rose cut flowers, any increasing effect by AsA on vase life was not noticed<sup>7-9</sup>. The treatment containing the combination of 100 mg L<sup>-1</sup> AsA + 200 mg L<sup>-1</sup> 8-HQS + 30 g L<sup>-1</sup> sucrose resulted in the significantly longest vase life, largest flower diameter, more dry and fresh weight and the highest mean uptake of preservative solution. The similar treatment without AsA had similar effect on all mentioned parameters but in addition, it had significantly lower anthocyanin content. A significant correlation between mean uptake of preservative solution and vase life and flower diameter was noticed as reported earlier<sup>17</sup> (*Table 2*).

We conclude that AsA could improve the anthocyanin content and flower diameter of flowers and adding it to preservative mixtures based on 8-HQS could improve its effect in a limited extent. Based on the increasing effect of 8-HQS on dry weight it seems that it may have supplied the cut flower as a carbon source as well as having germistatic effect in preservative mixture.

	Vase Life (days)	Dry weight (g)	Anthocyanin content (A530/657; mg FW)	Fresh weight - day 7 (g)	Flower diameter - day 7 (mm)	Stem diameter - day 7 (mm)
dry weight (g)	0.100					
Anthocyanin content						
(A530/657; mg FW)	0.632 **	-0.397 *				
fresh weight -day 7 (g)	0.396 *	0.627 **	0.086			
flower diameter - day 7 (mm)	0.578 **	0.590 **	0.287	0.849 **		
stem diameter - day 7 (mm) Mean uptake of preservative	0.240	0.248	0.138	0.594 **	0.563 **	
solution; ml	0.627 **	0.529 **	0.207	0.779 **	0.786 **	0.355 *

# Table 2: Cross correlation table

\*\*. Correlation is significant at the 0.01 level (2-tailed).

\*. Correlation is significant at the 0.05 level (2-tailed).

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